Lesson Plan

Course: MBMG 501 (Genetic Engineering)

Lecture: Analysis of Protein Structure

Lecturer/Instructor: Duangrudee Tanramluk, Ph.D.

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Academic Program: M.Sc. program in Molecular Genetics and Genetic Engineering and Ph.D. program in Molecular Genetics and Genetic Engineering

Date/Time: 9 November 2017 (1.30 pm – 3.30 pm)

Room: C405

Learning Objectives:

At the end of this lecture, student should be able to

1. Understand the amino acid structures and their properties

2. Describe the details of protein structure from various visualization styles

3. Describe information that can be derived from a protein structure model

Content Outlines:

- 1. Introduction to protein structure
 - 1.1 Hierarchical organization and size of proteins
 - 1.2 The 20 amino acid sidechains
- 2. Protein structure representation
 - 2.1 PDB file format
 - 2.2 Structure representation styles
- 3. Techniques for protein structure determination
 - 3.1 X-ray crystallography
 - 3.2 Cryo-Electron Microscopy
 - 3.3 Nuclear Magnetic Resonance
- 4. Information that can be derived from protein structure
 - 4.1 Example of structures and their usefulness
 - 4.2 Information derived from a PDB file of protein structure.

References:

- 1. Introduction to Protein Structure by Carl Branden & John Tooze
- **2.** Crystallography made crystal clear by Gale Rhodes (Chapter 2: An Overview of Protein Crystallography)
- **3.** Saibil, H. R., Macromolecular structure determination by cryo-electron microscopy. *Acta Cryst.* (2000), **D56**, 1215-1222.
- **4. Computational Structural Biology Methods and Applications** by Torsten Schwede and Manuel Peitsch (Chapter 27: Molecular Graphics in Structural Biology)
- **5. Structural Bioinformatics** by Philip E. Bourne and Helge Weissig (Chapter 2: Fundamentals of Protein Structure, Chapter 19: Inferring Protein Function from Structure)

Suggested Reading:

Subramaniam S, *et al.* Resolution advances in Cryo-EM enable application to drug discovery. *Curr Opin Struct Biol* (2016), **41:**194-202.

Learning Organization:

- 1. Lecture
- 2. Kahoot Quiz
- 3. Questions and answers

Learning materials:

- 1. Powerpoint slide
- 2. Handout
- 3. Internet/Website (https://ajarnwi.wordpress.com/courses/mbmg501/)

Student Assessment:

Written examination in colored answer sheets (2 questions)

Last modified: 24/10/2017

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Analysis of Protein Structure

Dr. Duangrudee Tanramluk

Contents:

- Introduction to protein structure
- Protein structure representation
- Techniques for protein structure determination
- Information that can be derived from protein structure

Objectives:

At the end of this lecture, student should be able to

- 1. Describe the details of protein structure from various visualization styles
- 2. Understand the principle of protein structure determination methods
- 3. Describe information that can be derived from a protein structure model.

Introduction

Proteins are essential biological molecules which play crucial roles in several cellular processes, e.g. catalyzing chemical reactions, transporting small molecules, serving as the sub-cellular structures, reacting against foreign bodies, and regulating other proteins in response to cellular environment.

The protein structure can be considered from several levels. The primary structure of a protein composes of amino acid sequence which constitutes a polypeptide chain. Some region of the polypeptide chain can form a local structure via series of hydrogen bonds from the repeating backbone torsion angles referred as the secondary structure. Combination of several of secondary structures gives rise to a three dimensional fold called the tertiary structure. Within the fold space, common structural motifs and domains can be recognised. For many proteins, a larger organization called the quaternary structure is required for the protein function. One can use the three dimensional structure of a protein to rationalize the molecular interaction underlying its biological process after obtaining the functional information from biochemical experiments. Several tools have been developed to visualize the three dimensional structures and assist in molecular design. With the advent of structural genomics, structural biologists are working on the structure of all the proteins that a genome could produce without knowing their function. Functional annotation becomes an important step toward characterization of cellular and physiological roles of gene products.

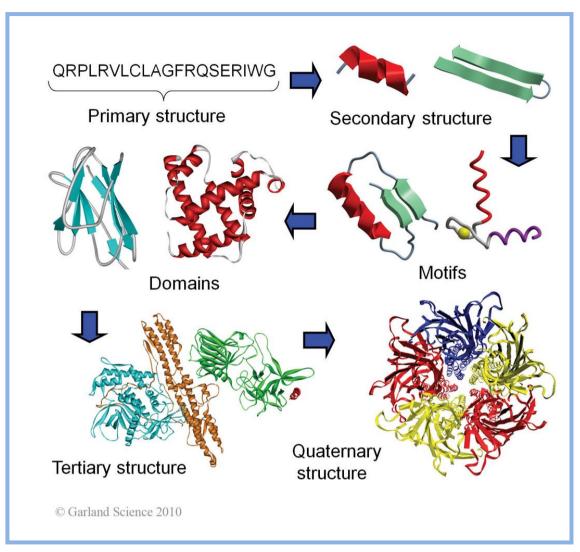


Figure 1 Protein structure hierarchy (taken from Bernhard Rupp, Biomolecular Crystallography: Principles, Practice, and Applications to Structural Biology (2010), Garland Science)

Protein Structure Representation

Upon publishing the protein structure in a scientific journal, structural biologist usually deposits the structural data in a repository called the Protein Data Bank (PDB: <u>http://www.pdb.org</u>). Each file in the PDB is given a 4-character identification number (PDB ID) and has to conform to a textual format called the PDB format. You can download the (.pdb) file and visualize molecules in a molecular viewer (e.g. Pymol, Discovery Studio). Figure 2 shows the ATOM record which is a section of the PDB file that most molecular viewers can read in order to draw atoms and bonds. Notice that we can hardly locate the position of hydrogen atom. Therefore, most of the structure model was built without hydrogen atoms and hence we usually omitted them in the PDB file.

ATOM/			me Chain		х	Y	Z	occupancy	
HETATM	Atom ID		Resname	Resnum	ber			B-factor	Element
+	1	+-	2	+	3+	-4+-	5+	+6+	-78-
ATOM	581	N	GLU E	86	2.988	5.846	34.351	1.00 31.46	N
ATOM	582	CA	GLU E	86	3.910	5.294	33.377	1.00 27.74	С
ATOM	583	С	GLU E	86	4.552	6.423	32.576	1.00 29.59	С
ATOM	584	0	GLU E	86	5.756	6.448	32.362	1.00 25.21	0
ATOM	585	CB	GLU E	86	3.174	4.346	32.429	1.00 30.03	С
ATOM	586	CG	GLU E	86	3.108	2.900	32.912	1.00 30.92	С
ATOM	587	CD	GLU E	86	4.436	2.381	33.379	1.00 28.34	С
ATOM	588	OE1	GLU E	86	5.387	2.395	32.574	1.00 31.36	0
ATOM	589	OE2	GLU E	86	4.532	1.959	34.540	1.00 25.12	0
ATOM	590	N	HIS E	87	3.729	7.348	32.110	1.00 31.69	N
ATOM	591	CA	HIS E	87	4.222	8.472	31.348	1.00 33.42	С
ATOM	592	С	HIS E	87	5.113	9.344	32.215	1.00 33.67	С
ATOM	593	0	HIS E	87	6.125	9.867	31.772	1.00 37.08	0
ATOM	594	CB	HIS E	87	3.053	9.292	30.875	1.00 35.84	С
ATOM	595	CG	HIS E	87	2.139	8.547	29.970	1.00 37.98	С
ATOM	596	ND1	HIS E	87	0.802	8.365	30.250	1.00 40.85	N
ATOM	597	CD2	HIS E	87	2.354	7.979	28.761	1.00 38.02	с
ATOM	598	CE1	HIS E	87	0.233	7.718	29.247	1.00 41.67	с
ATOM	599	NE2	HIS E	87	1.153	7.473	28.331	1.00 35.98	N
ATOM	600	N	THR E	88	4.719	9.510	33.463	1.00 31.68	N
ATOM	601	CA	THR E	88	5.484	10.331	34.367	1.00 32.97	С

Figure 2 Some part of the ATOM record in the PDB ID: 1STC. This PDB file 1stc.pdb was opened with text editor EditPlus to show the atomic coordinates of glutamate (GLU), atom ID 581-589 from residue number 86 of chain E, and histidine (HIS), atom ID 590-599 from residue number 87.

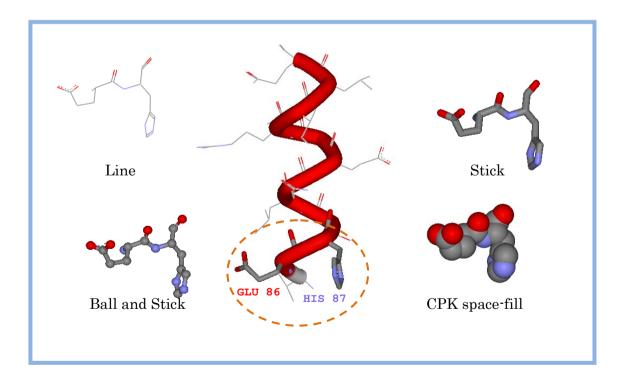


Figure 3 Atomic representations of the residue number 86 and 87 which are parts of a helix from the PDB ID: 1STC. The peptide backbone is displayed as a large spiral tube, portraying its helical character. (Figures created using Discovery Studio package).

When coloring atom by element type, carbon atoms are usually colored in grey, oxygen atoms are in red, nitrogen atoms are in blue, and sulphur atoms are in yellow. The Corey-Pauling Koltun (CPK) model uses this coloring scheme and displays each atom as a sphere. Coloring atoms by their electrostatic charge is useful for visualizing binding interaction. For instance, glutamate and aspartate can form similar interaction due to their partial negative charge characters, so we can represent them as a part of the molecular surface with the same color.

Table 1 The 20 amino acid side chains in stick representation. The three-letter amino acidname is on the top with one-letter abbreviations in parentheses. Note that these side chainscan adopt several conformations. (Figures created using Discovery Studio package).

GLY (G)*	ALA (A)	SER (S)	CYS (C)
CA CA	Ch Ch	CBG	СВ
VAL (V)	LEU (L)	ILE (I)	THR (T)
31 0B 06 052	СБ 5D1	GD1 G2	CB GI
ASP (D)	GLU (E)	ASN (N)	GLN (Q)
	CD CD E2	02 05 01	
MET (M)	LYS (K)	ARG (R)	PRO (P)
GD SE	CB CD CE	B CO CO NE CO HH	C C C C C C C C C C C C C C C C C C C
HIS (H)	PHE (F)	TYR (Y)	TRP (W)
B CD2	CDT CE1	CD2 CZ OH CD1 CE1	NE1 CE2 CH2 CD1 CG2 CZ3 CG3 CG2 CE3

(*Glycine has no side chain, so its main chain is shown instead)

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When viewing a protein structure by showing every atoms and bonds, we can hardly see the polypeptide chain that those atoms belong to. Interactions among different chains can be observed by coloring carbon atoms from each polypeptide chain as one color per one chain. In order to see the organization of secondary structure elements, alpha helices are shown with spiral shape ribbons and beta strands are shown with thick arrows. Schematic styles can show the polypeptide chain direction from the head of the cylinder rods and the arrows. Some side chains that involve in making interactions and also the ligand can be shown with stick model and the rest of the side chains are usually omitted for clarity.

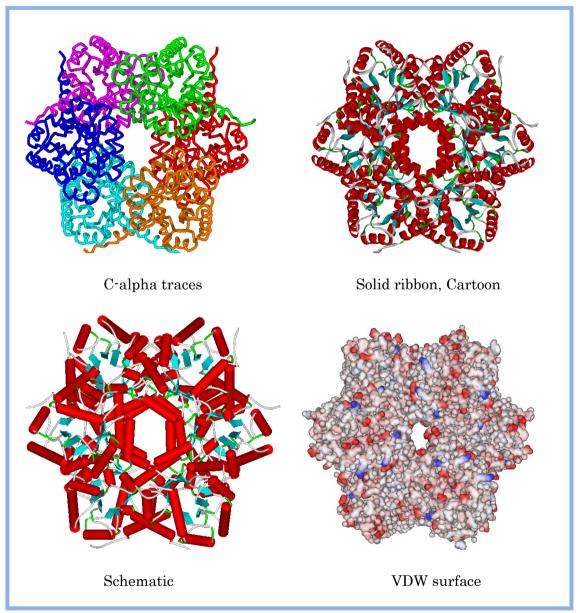


Figure 4 Amino acid representation of the PDB ID: 2P10. In the schematic style, alpha helices are displayed as rods, beta strands are represented by arrows, and turns are drawn as traces. In the VDW surface style, each atom is represented as a sphere with its van der Waals radius. (Figures created using Discovery Studio package).

Techniques for Protein Structure Determination

Various techniques can be used to determine the three dimensional structure of a protein. The majority of structural data are obtained from X-ray diffraction, nuclear magnetic resonance spectroscopy (NMR), or cryo-electron microscopy (cryo-EM) experiments. The choice of the method depends on several considerations. For example, the concentration of protein sample for both NMR and X-ray diffraction should be at least 1 mg/ml, while the protein sample for cryo-EM is more diluted and hence require a much smaller amount of sample. X-ray diffraction gives more precise value of atomic coordinates and its molecular size can range from a very small molecule to mega-daltons of protein complex. However, not all protein can be crystallized and only some proteins can diffract X-ray. Structures obtained from cryo-EM are usually in the near native physiological form and the molecule can be very large, but it often has poor resolution. Some of the strengths and weaknesses are summarized in Table 1.

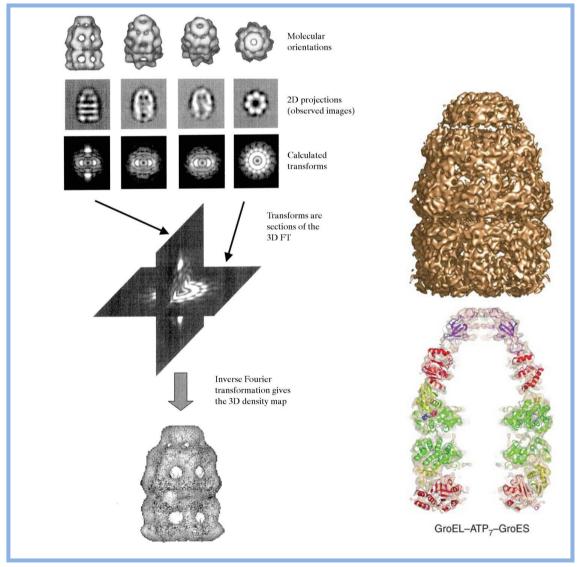


Figure 5 Obtaining a cryo-EM structure (taken from Saibil, H. R., *Acta Cryst.* (2000). D56, 1215-1222 and Ranson N. A., et al. *Nat Struct Mol Biol* (2006), 13, 147 – 152)

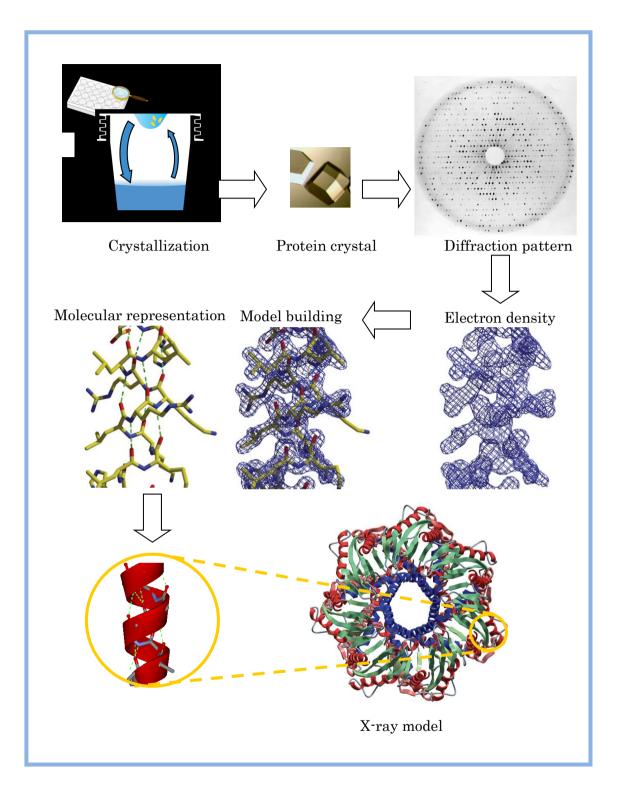
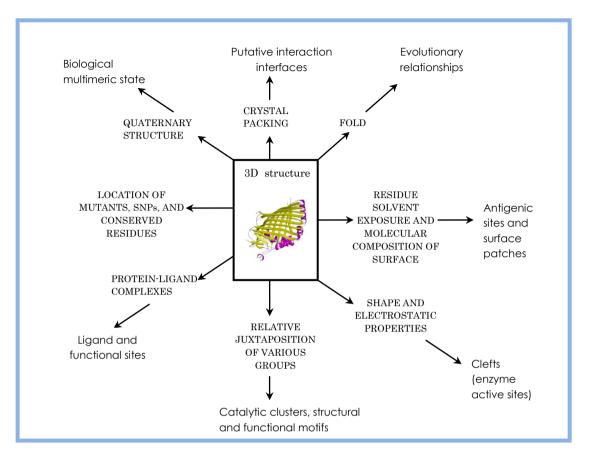


Figure 6 X-ray crystallography (pictures were adapted from Bernhard Rupp, **Biomolecular Crystallography: Principles, Practice, and Applications to Structural Biology** (2010), Garland Science and content of the figures were created using Discovery Studio package).

Table 2 Advantages and limitations of protein structure determination methods

Factor	X-ray	NMR	Cryo-EM
Sample	Crystal (from ~2 to 10 mg/ml homogenous protein)	Homogenous, monodisperse solution (~0.2 mM)	Thin cross section (<2,000 Å), crystals, ordered assemblies or isolated particles usually in native, hydrated state.
Size	Mega-dalton (MDa) range is possible	Usually < 40 kDa, up to 1 MDa in rare cases	More than 300 kDa
Data collection	Diffraction pattern	Peaks that can be interpreted to restraints	Low contrast images
Resolution	0.5 - 5.0 Å (median ~2.0 Å)	-	Usually >3.0 Å, about 10 Å is normal. An extremely rare case is at < 2.0 Å resolution.
Conformation	As in crystal lattice	Dynamic processes	Can also trap transient state by rapid freezing
Final structure	One model of atomic coordinates with B-factor	An ensemble of atomic coordinates	EM electron density map, Cα trace.



Information that can be derived from 3D structure

Figure 5 From structure to function: A summary of information that can be derived from 3D structure relating to biological function (reproduced from Philip E. Bourne and Helge Weissig, **Structural Bioinformatics** (2003) John Wiley & Sons, Inc.)

If you have a PDB identification number (PDB ID), a protein name, or its amino acid sequence, you can search for a PDB ID of the structure from various websites. In this lecture, I will show you how to use a website called **PDBsum** in order to start exploring your structure. It provides several links to visualize the structure. To display a structure from PDBsum on a Java-enabled web browser, you can just activate the program **Jmol** on the left of Figure 7 or Figure 9. The URL for PDBsum is:

http://www.ebi.ac.uk/pdbsum

For each PDB code, this server extracts most of the information the can be found before the ATOM record of a PDB file.

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For example:

• Title

The name of the protein with ligand and/or nucleic acid in the complex is shown.

• Chains

It can identify all the polypeptide chains that are associated with the structure. The chain length (number of amino acids in the chain) is also provided.

• Details about the protein and the source organism

The *Universal prot*ein resource (Uniprot) is the gold standard for annotation of amino acid sequence. The UniprotKB entry is in an abbreviation form of protein_organism. If you enter PDB ID: 1B39 in PDBsum, you will obtain Uniprot ID: CDK2 HUMAN which is for the cell division protein kinase 2 from human.

• Resolution

It is the uncertainty of atomic position in a model. If the value, in angstrom (Å), is smaller, the electron density from the map is more accurate. Cryo-EM density map cannot be used for analyzing atomic interaction because its resolution is so poor that it cannot accurately define the inter-atomic distance in the model. A quick look at resolution, i.e. high resolution is < 2 Å, can imply the good model quality.

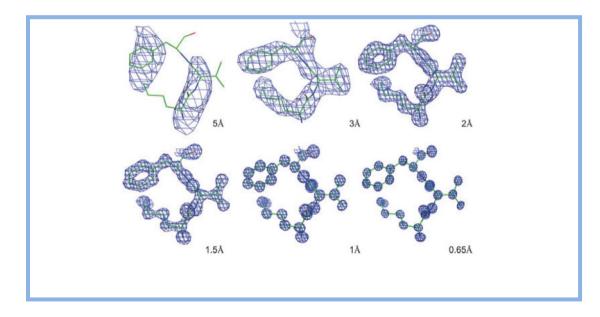


Figure 6 Effects of resolution. The electron density map (mesh) obtained at 0.65Å resolution agrees with the position of atoms in the line model better than the one obtained at 5Å resolution. (Taken from Wlodawer, A., et al. *FEBS J.* (2008), 275, 1–21.)

• Bound ligand and metal ion

In the PDB file, a heterogen is a molecule, a prosthetic group, a solvent, or an ion that is not a part of the protein and its atoms are listed in the HETATM section of the PDB file. A small molecule is usually identified with its heterogen three-letter code, while an ion is normally identified by its element symbol.

• Authors, references, and abstract of the published literature

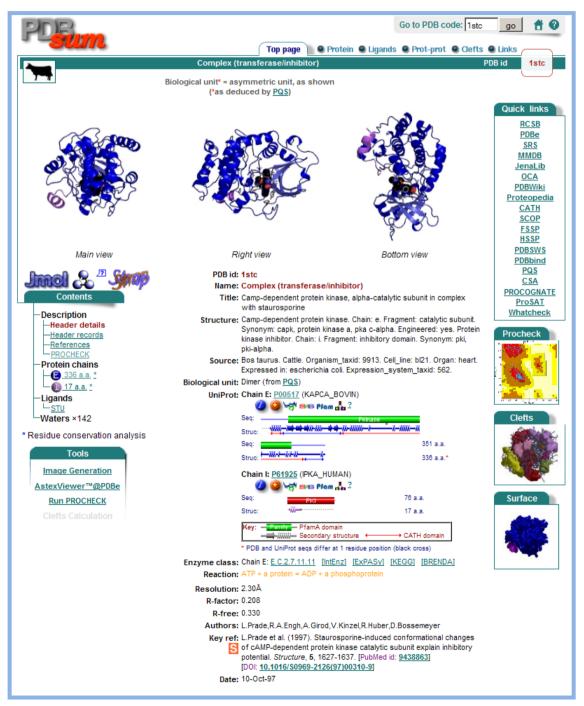


Figure 7 PDBsum screenshot upon searching for PDB ID: 1STC from www.ebi.ac.uk/pdbsum.

In addition to the information that can be found in the head of the PDB file, there are also some 'derived' data that are included in PDBsum. For example:

• Protein-ligand interactions

In order to bind protein well, the ligand usually has complementary electrostatic potentials with the amino acids in the protein binding site. Hydrophobic interactions can also govern protein packing and the hydrogen bonding interactions are important for ligand binding.

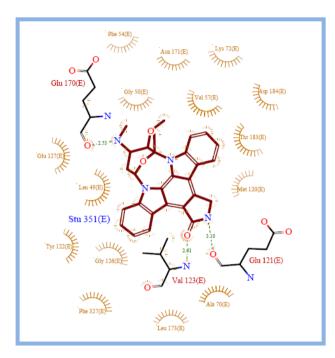


Figure 8 Ligand interactions obtained from the 'Ligands' tab from the top of Figure 7. The program LIGPLOT shows called ligand staurosporine а (heterogen name:STU) interacts with various amino acids in the pocket of the cAMP-dependent protein kinase (PDB ID: 1STC). Hydrogen bonds are shown with the dash lines. To see more detailed analysis of the ligand interaction with series of proteins, the server MANORAA.org can be used.

• Protein-protein interactions

There is a 'Prot-prot' tab to show interacting interfaces between this protein and other proteins on the top of Figure 7.

• Cleft positions

The largest cleft is a possible active site and may be useful for inhibitor design.

• Ramachandran plot

PROCHECK is the underlying program that calculate Ramachandran plot for PDBsum. You can assess whether the structure has a bad model quality by observing the phi-psi angles whether they are distributed in the usual location. There are usual locations where alpha helices, beta sheets and left handed helices occurred with glycine scattered around. If these angles deviated too much from the standard locations, it may mean that the structure was not correctly built or was fabricated.

	Chain 😉 (336 residues)				
	UniProt code:P00517 (KAPCA_BOVIN) [Pfam]				
A 10 55	CATH structural classification (2 domains) :				
2 R R R	Domain Links CATH no. Class Architecture 1 CATH Lass Architecture				
ON STATES	2 CATH 🚠 3.30.200.20 = Alpha Beta 2-Layer Sandwich				
▼ <i>∨</i>	Η1 Η2 Α β Α ββ Α				
Protein chain E highlighted	VKEFLAKAKEDFLKKWENPAQNTAHLDOFERIKTLGTGSFGRVMLVKHMETGNHYAMKI 15 20 25 30 35 40 45 50 55 60 65 70				
(click to view)	H3 H4 A A H5				
mol 🏡 💥 柳	· · · · · · · · · · · · · · · · · · ·				
Motifs	LKQKVVKLKQ I EHTLNEKR I LQAVNF PFL VKLEF SFKDNSNL YMVME V VPGGEMF SHLR				
-Secondary structure 	7's 8'o 8's 9'o 9's 10'o 10's 11'o 11's 12'o 12's 13'o				
Residue conservation ProMotif					
- <u>3 sheets</u>					
— <u>4 beta hairpins</u> — <u>5 beta bulges</u>	IGRFSEPHARFYAAQIVLTFEYLHSLDLIYRDLGPDNLLIDQQGYIQVTDFGFAKRVKG 135 140 145 150 155 160 165 170 175 180 185 190				
— <u>9 strands</u> — <u>16 helices</u>	H3 H9 B H10				
— <u>20 helix-helix interacs</u> —22 beta turns	<u> </u>				
1 gamma turn					
-Catalytic residues	19'S 20'O 20'S 21'O 21'S 22'O 22'S 23'O 23'S 24'O 24'S 25'O H11 0 H12 0 0 H13 H14 H15				
— <u>D166-K168</u> — <u>D166-K168-T201</u>					
<u>N171-D166-K168</u> PROSITE	VRFPSHFSSDLKDLLRNLLQVDLTKRFGNLKNGVNDIKNHKWFATTDWIAI VQRKVEAP				
PROTEIN KINAS	255 260 265 270 275 280 285 290 295 300 305 310				
PROTEIN KINAS	ββββ				
<u>e st</u>	IPKFKGPGDTSNFDDYEEEE IRVS INEKCGKEF SEF				
Protein	315 320 325 330 335 340 345 350				
-Chain 💽 336 a.a. - <u>Chain 🕕 17 a.a.</u>	Key:				
	Sec. struct All-Control Helices labelled H1, H2, and strands by their sheets				
	A, B, Helix Strand				

Figure 9 By clicking on the protein chain, one can obtain the amino acid sequence, the sequence diagram and the CATH identification number. The protein class and the architecture of this chain can be recognized from the CATH ID. This picture also shows residues that are in contact with the ligand (dots) and active site residues (tiny rectangles). Secondary structures are shown on top of sequence (cartoon). Source: www.ebi.ac.uk/pdbsum.

• Fold classification

CATH (*C*lass, *A*rchitecture, *T*opology, *H*omologous superfamily) is a server for protein fold classification. PDBsum provides a link to CATH from the 'Protein' tab. According to CATH, a protein can be primarily classified as mainly alpha, mainly beta, mixed alpha-beta, or few secondary structures. If two proteins have similar sequences and adopt the same fold, they can be evolutionary related. With a CATH ID, you can look for structures that have the same protein fold with your queried PDB ID.

• Enzyme Commission number (E.C.)

Every enzyme code can be identified with the letter "EC" followed by four numbers separated by periods. If your protein is an enzyme, it will have its own E.C. number to identify that it is an oxidoreductase, transferase, hydrolase, lyase, isomerase, or ligase.

• Pathways

The Kyoto Encyclopedia of Genes and Genomes (KEGG) is one of the most comprehensive interaction pathway resources. A pathway can be retrieved if your protein is known to take part in metabolism, genetic information processing, environmental information processing, cellular processes, organismal systems, or human diseases. This is how you can discover the role of your protein in relation with other proteins in the pathway.

This reading material introduced the concepts of the protein structure representation, the differences among major structure determination methods, and also an overview of structural information that can be obtained from the PDB file. These key points and details of the figures can provide students with background knowledge to understand literatures on structural biology and the relating fields.